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With amended claims.

TREATING CANCER WITH ENTEROTOXIN
FROM USE OF STAPHYLOCOCCUS AUREUS
ENTEROTOXINS IN TREATMENT
OF CANCER + MAX. 30
ADMINISTERED BY INJECTION,
AND HAVE SAME TUMORICIDAL
ACTIVITY

(54) Title: TUMOR KILLING EFFECTS OF ENTEROTOXINS AND RELATED COMPOUNDS

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10      20      30      40      50
STR-PPPSQLOKSHLVKTKYIIFKQVTL-----VTKDNVSVQDLSHDLIYKVS--
111 111 111 111 111 111 111 111 111 111
120      130      140      150
LSQDPDPKDFELKSS--K-PTGLKDKCKV-LYNDKVSALNKSINLTF--DLIYLSIK
10      20      30      40      50
-----60      70      80      90-----ENHAKSAC
111 111 111 111 111 111 111 111 111 111
100      110      120      130      140      150
LYGGVTNHGKMLIPK-----IVVKVSIIDGIGSLFDIEQINQNCGRIS--YTVKYL
111 111 111 111 111 111 111 111 111 111
120      130      140      150      160
HYGGVTENGNNQLD--KYIASITVKTEDGKMLSTFVOTNKKVTAQGLDYLTRYLV
120      130      140      150      160
170      180      190      200
DNKQLYTNQPSKYITGYIKFIPKQKESFVDFPFPEL--PTQSKYLMKDKNITLDSNTS
11 11 11 11 11 11 11 11 11 11
170      180      190      200      210      220
KXKLYEYNHSPYITGYIKFII-NEHSFVYDQFAPGNKFDQSKYLPQYNDKQVDSKDV
170      180      190      200      210      220
220
QIEVYLTK
111111
KIEVYLTKKK
220

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AS
STAPHYLOCOCCUS
PROTEIN A
WITHOUT
POTENTIAL
TOXIC
SIDE REACTIONS

(57) Abstract

Staphylococcal enterotoxins obtained by secretion from Staphylococcus aureus, by expression of enterotoxins in other bacteria or cells, or by chemical mutagenic treatment of S. aureus strains are used in treatment of cancer as tumoricidal agents. Enterotoxins (A, B, C, D, E and F) can be administered via simple intravenous injection at dosages between 0.5 µg/kg body weight to 150 µg/kg. Non-steroidal, anti-inflammatory agents and prostaglandin synthesis inhibitors such as ibuprofen may be simultaneously administered to attenuate toxic reactions from the enterotoxins. Streptococcus pyrogenic exotoxin and alpha hemolysin, which have structural and functional homology to the enterotoxins, are also useful in tumoricidal treatment. The enterotoxin gene transfected into tumor cells resulting in a tumor cell with surface expression of the minor lymphocyte stimulating locus with consequent potent activation and proliferation of T lymphocytes especially those with V_H specificity, would also be useful in this application.

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Tumor Killing Effects
Of Enterotoxins and Related Compounds

Related Application Data

This application is a continuation-in-part application of Application Serial No. 07/466,577, filed on January 17, 1990, which is a continuation-in-part application of Application Serial No. 07/416,530, filed October 3, 1989.

Technical Field

This invention relates generally to tumoricidal compositions and methods, and more specifically to the use of enterotoxins derived from, or homologous to those derived from Staphylococcus aureus, to the use of streptococcal pyrogenic exotoxins, TSST-1, mycoplasma arthritidis, Mis antigens, homologous synthetic polypeptides and derivatized enterotoxins.

Background Of The Invention

Therapy of the neoplastic diseases has largely involved the use of chemotherapeutic agents, radiation and surgery. However, results with these measures, while beneficial in some tumors, has had only marginal or no effect in many others, while demonstrating unacceptable toxicity. Hence, there has been a quest for newer modalities to treat neoplastic diseases.

In 1980, tumoricidal effects were demonstrated in four of five patients with advanced breast cancer utilizing therapy with plasma perfused over Staphylococcal Protein A. Terman, D.S., Young, J.B., Shearer, W.T., Ayus, C., Lehane, D., Mattioli, C., Espada, R., Howell, J.F., Yamamoto, T., Zaleski, H.E., Miller, L., Frommer, P., Feldman, L., Henry, J.F., Tillquist, R., Cook, G., Daskal, Y., New Eng. J. Med., 305, 1195, 1981. This elaborate system

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involved the administration of patient plasma which was perfused over a solid surface to which Staphylococcal Protein A was chemically attached. Protein A was prepared by batch fermentation of Staphylococcus. It was isolated from the media and partially purified by affinity chromatography.

While the initial observations of tumor killing effects with the immobilized Protein A perfusion system have been confirmed, additional results have been inconsistent. The explanation of these inconsistencies appears to be as follows. First, commercial Protein A has been shown to be an impure preparation, as evident from polyacrylamide gel electrophoresis studies. Secondly, various methods of the immobilization of Protein A to solid supports have been used, sometimes resulting in loss of biological activity of the plasma perfusion system. Thirdly, the plasma used for perfusion over the immobilized Protein A has been stored and treated in different ways, sometimes resulting in inactivation of the system. Moreover, the antitumor element present in this extremely complex perfusion system has not been previously defined. The system contained an enormous number of biologically active materials, to include Staphylococcal Protein A itself, Staphylococcal proteases, nucleases, exotoxins, enterotoxins and leukocidin, as well as the solid support and coating materials. Additional products included several anaphylatoxins generated in plasma after contact with immobilized Protein A. Finally, it is speculated that the biological activity of the system was due to extraction by Protein A of immunosuppressive immune complexes capable of blocking the host's antitumor response.

The present invention demonstrates that isolated Staphylococcal enterotoxins identified initially as

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contaminants in commercial Protein A can reliably reproduce the tumoricidal reactions and toxicity observed with the whole perfusion system. As such, these materials appear to represent the most active tumoricidal components in the Protein A perfusion system matrix. These materials demonstrate tumoricidal activity in small doses and produce tumoricidal effects and toxicity identical to that observed in animals and man with the whole Protein A perfusion system. However, the tumoricidal effects may be produced by a simple intravenous injection. Therefore, it has been possible to completely eliminate the elaborate and complex Protein A perfusion system, with its enormous number of component parts, unpredictable performance and potential toxicity problems. This system may be replaced by the enterotoxins which may be administered via simple intravenous injection and have the distinct advantages of convenience, reliability and safety over the cumbersome, inefficient and often ineffective extracorporeal Protein A perfusion system. There is no prior report in the literature or elsewhere of antitumor effects ascribable to this group of proteins.

Enterotoxins have distinct advantages in inducing tumor killing effects over the more cumbersome and elaborate Staphylococcal Protein A plasma perfusion systems. One advantage is that enterotoxins are relatively simple proteins that may be infused after being solubilized in saline. This solubility obviates the need to immobilize Protein A or other biologicals on a solid support, and eliminates the requirement for perfusing plasma over a solid surface. Moreover, it bypasses problems associated with potential toxic reactions to impurities of Protein A. Consequently, enterotoxins

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appear to be far safer and more effective than previously described systems. Moreover, the system requires no elaborate sterilization and there is no problem with potential leaching of immobilized materials or chemical products from an inert surface as there would be with an extracorporeal column. Hence, this product offers decided advantages of effectiveness and convenience over the original system. Indeed, all evidence points to enterotoxins as being the most active antitumor product in the Staphylococcal Protein A plasma perfusion system.

Summary Of The Invention

The present invention provides enterotoxins derived from Staphylococcus aureus which are useful by themselves for the treatment of cancer. Enterotoxins are known to have molecular weights ranging from 22,000 to 38,000. They are heat stable, and resistant to trypsin digestion. According to one aspect of the present invention, enterotoxins isolated from media which is supporting the growth of various Staphylococcus aureus organisms are used in relatively pure form. When administered to subjects having tumors, the preparation induces a tumoricidal reaction: a hemorrhagic necrosis in tumor resulting in tumor regression. It should be understood that the term, "tumoricidal reaction," as used herein, means that the material under discussion promotes or assists in the killing of tumor cells.

Streptococcal pyrogenic exotoxin A, which has been shown to have statistically significant structural homology to enterotoxin B, is also useful for the treatment of cancer. The structural homology is supported by similar clinical responses: hypotension, fever, chills and septic shock. Additional biological properties common to these

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materials include mitogenic effects, interferon, interleukin and tumor necrosis factor induction activity.

Synthetic polypeptides with substantial structural homology and with statistically significant sequence homology and similarity to enterotoxin A, enterotoxin B and streptococcal pyrogenic exotoxins, including alignment of cysteine residues and similar hydropathy profiles, are also described as effective in tumoricidal therapy.

Brief Description of Figures

Figure 1 shows the alignment of amino acid sequences of staphylococcal enterotoxins and their relatives.

Figure 2 shows the alignment of amino acid sequences of mature streptococcal pyrogenic exotoxin A and staphylococcus aureus enterotoxin B.

Figure 3 shows the hypothetical structure for the complex of Class II MHC, T cell receptor, and staphylococcal enterotoxins or MIs.

Description Of The Specific Embodiments

The enterotoxins of Staphylococcus aureus form a group of serologically distinct extracellular proteins, designated A, B, C₁, C₂, C₃, D, E and F. These proteins are recognized as the causative agents of Staphylococcal food poisoning. Enterotoxin F appears to be important in the pathogenesis of the Staphylococcal toxic shock syndrome. Ingestion of preformed enterotoxin in contaminated food leads to the rapid development (within two to six hours) of symptoms of vomiting and diarrhea that are characteristic of Staphylococcal food poisoning.

The enterotoxin proteins are of similar molecular weight. Characteristically, they have a

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disulfide loop near the middle of the molecule, and are easily soluble in water and salt solutions. They are relatively resistant to proteolytic enzymes and to heat. The higher level structural similarities between the enterotoxins is in agreement with the clinical picture where all of the enterotoxins seem to produce similar effects of sepsis, hypotension and fever. General properties of the enterotoxins are given in Table 1.

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TABLE 1A
SOME PROPERTIES OF THE ENTEROTOXINS

	Enterotoxin				
	A ^a	B ^b	C ^c	D ^d	E ^e
Emetic dose (ED ₅₀)(g/monkey)	5	5	5	5-10	
Nitrogen content (%)	16.5	16.1	16.2	16.0	
Sedimentation coefficient (S _{20,w})(S)	3.04	2.78	3.00	2.90	
Diffusion coefficient (D _{20,w})(x 10 ⁻⁷ cm ² sec ⁻¹)	7.04	8.22	8.10	8.10	
Reduced viscosity (ml/gm)	4.07	3.81	3.4	3.7	
Molecular weight	34,700	30,000	34,100	34,000	
Partial specific volume	0.726	0.726	0.728	0.725	
Isoelectric point	6.8	6.6	6.6	7.0	
Maximum absorption (mμ)	277	277	277	277	
Extinction (E _{1%} ^{1cm})	14.3	14.4	12.1	12.1	

^a F.S., Thachani, K., Schantz, E.J., Bergdoll, M.S., *Biochemistry* 5, 3201, 1966.

^b Bergdoll, M.S., Borja, C.R., Avena, R.M., *J. Bacteriol.* 90, 1461, 1965.

^c Borja, C.R., Bergdoll, M.S., *Biochemistry* 6, 1437, 1967.

^d Avena, R.M., Bergdoll, M.S., *Biochemistry* 6, 1474, 1967.

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TABLE 1B
Physicochemical Properties of Staphylococcal Enterotoxins^a

Property	Enterotoxin				
	A ^a	B ^b	C ^c	D ^d	E ^e
Emetic dose for monkey (g)	5	5	5	5-10	
Sedimentation coefficient (S _{20,w})	3.03	2.89	3.0	2.9	
Molecular weight	27,800	28,366 ^f	28,000	34,100	27,300
Isoelectric point	7.26	8.6	8.6	7.0	7.4
C-terminal residue	Serine	Lysine	glycine	lysine	Threonine
N-terminal residue	Alanine	Glutamic acid	Glutamic acid	Glutamic acid	Serine

^a Schantz, E.J., Rossler, W.G., Woodburn, M.J., Lynch, J.H., Jacoby, H.H., Silverman, S.J., Gorman, S.J., *Biochemistry* 11, 340, 1972.

^b Schantz, E.J., Rossler, W.G., Wagan, J., Spero, L., Bunney, D.A., Bergdoll, M.S., *Biochemistry* 4, 1011, 1965.

^c Borja, C.R., Bergdoll, M.S., *Biochemistry* 6, 1437 (1967).

^d Avena, R.M., Bergdoll, M.S., *Biochemistry* 6, 1474 (1967).

^e Chung, P.C., Bergdoll, M.S., *Biochemistry* 10, 1937, 1970.

^f Borja, C.R., Fanning, E., Huang, I.T., Bergdoll, M.S., *J. Biol. Chem.* 247, 2456, 1972.

^g Dayhoff, M. ed. (1972) Data Section. In *Atlas Protein Sequence Structure* 5 :

0227 National Biomedical Research Foundation, Washington, D.C. (determined from the amino acid sequence of Huang and Bergdoll, 1970). Huang, I.T., Bergdoll, M.S., *J. Biol. Chem.* 245, 3493, 1970.

^a Modified from Bergdoll, M.S., Crisp, J.E., Gould, S.S., Enterotoxin Synthesis by the Staphylococci. In: *Recent Advances in Staphylococcal Research*, pp. 307-316, Tait, V.V. (Ed.) Ann. N.Y. Acad. Sci. Vol. 236.

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Amino acid compositions of enterotoxins A, B, C₁, C₂ and E reveal a high content of lysine, aspartic acid and tyrosine. Enterotoxins A and E are similar in methionine, leucine and arginine content, differing in this regard from enterotoxins B, C₁ and C₂. The amino acid sequence of enterotoxin B was found to consist of 239 amino acids. Half-cystine residues found at positions 92 and 112 form a disulfide bridge, and it has been suggested that the primary structure in this region may be common to all of the enterotoxins.

The protein sequences and immunological cross reactivity of the enterotoxins reveal that they can be divided into two related groups. SEA

(Staphylococcal enterotoxin A), SEE and SED constitute one group, and SEB, SEC and Streptococcal pyrogenic exotoxin A (SPEA) make up the second group. Amino acid sequences show that SEA and SEE are almost identical and that SEB, SEC and SPEA share regions of similar sequence. SED is moderately related to both groups although it is more similar to the SEA group.

There is a striking amino acid similarity among enterotoxins A, B, C, D and E in the region immediately downstream from cysteine located at residue 106 in SEA. A second region at residue 147 also shows a highly conserved sequence. These regions are contained on the peptide fragment of SEC, shown to contain the active sites for emesis and diarrhea. The mitogenic region resides in the C terminal tryptic fragment of SEC, implying that other regions of sequence similarity exist. Amino acid sequence similarities and congruences are given in Tables 2-4.

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TABLE 2*
SEQUENCE SIMILARITIES AMONG THE
PYROGENIC TOXINS AND ENTEROTOXINS

TOXIN	SEQUENCE		
	106	119	147
SEA	CHYGGVTLHDNRRL		KKNVTQELDLQARRYL
SEB	CHYGGVTEHGHOL		KKNVTQELDYLTRYL
SEC1	CHYGGITTEGHNIF		KKSVTQELDIKARNPL
SED	CTYGGVTFEGNKL		KKNVTQELDAQARRYL
SEE	CHYGGVTLHDNRRL		KKEVTQELDLQARRYL
SPEA	CTYGGVTNHEGHEL		KKNVTQELDYKRRYL
Consensus	CHYGGVTLHEGNL		KKNVTQELDLQARRYL
TSST-1	INPQISGVNTTKL		KKQLAISTLDPEIRHQL

* Landolo, J.J., Annu. Rev. Microbiol., 43, 375, 1989.

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TABLE 3

Amino Acid Composition of the Enterotoxins
(g 100g protein)

Amino Acid	Enterotoxin				
	A ^a	B [†]	C [‡]	C [§]	E [¶]
Lysine	11.26	14.85	14.43	13.99	10.83
Histidine	3.16	2.34	2.91	2.87	3.04
Arginine	4.02	2.69	1.71	1.75	4.50
Aspartic acid	15.53	18.13	17.85	18.38	15.10
Threonine	5.96	4.50	5.31	5.80	6.36
Serine	2.99	4.05	4.58	4.81	4.72
Glutamic acid	12.36	9.45	8.95	8.93	12.15
Proline	1.35	2.11	2.16	2.23	1.93
Glycine	2.96	1.78	2.99	2.90	4.10
Alanine	1.94	1.32	1.85	1.61	2.38
Half-cystine	0.66	0.68	0.79	0.74	0.81
Valine	4.93	5.66	6.50	5.87	4.36
Methionine	0.96	3.52	3.20	3.60	0.45
Isoleucine	4.11	3.53	4.09	4.02	4.30
Leucine	9.78	6.86	6.54	6.13	10.08
Tyrosine	10.63	11.50	9.80	10.27	9.79
Phenylalanine	4.31	6.23	5.35	5.25	4.47
Tryptophan	1.46	0.95	0.99	0.84	1.51
Amide NH ₂	1.80	1.66	1.71	1.62	1.66
TOTAL	98.37	100.15	100.00	99.99	100.88

^a Schantz et al., 1972.

[†] Bergdoll, M.S., Chu, F.S., Huang, I.Y., Rowe, C., Shih, T., Arch Biochem Biophys, 112, 104, 1965.

[‡] Huang, I.Y., Shih, T., Borja, C.R., Avena, R.M., Bergdoll, M.S., Biochemistry, 6, 1480, 1967.

[§] Borja et al., 1972.

[¶] From Bergdoll, M.S., Huang, I.Y., Schantz, E.J., J. Agric. Food Chem. 22, 9, 1974.

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TABLE 4¹

Amino Acid Compositions of TSST-1a and 1b^a

Amino acid	Amino acid composition	
	TSST-1a residues per mole ^b	TSST-1b residues per mole ^b
Aspartic acid	26	27
Threonine	21	20
Serine	20	20
Glutamic acid	20	20
Proline	10	8
Glycine	13	14
Alanine	4	5
Half-cystine	0	0
Valine	5	5
Methionine	0	0
Isoleucine	15	15
Leucine	14	16
Tyrosine	10	8
Phenylalanine	7	7
Histidine	5	5
Lysine	23	24
Tryptophan	ND ^c	ND ^c
Arginine	4	5
	197	199
		194

¹ Blomster-Hautamaa, D.A., Schillevert, P.M., Methods in Enzymology, 163, 37, 1988.

^a Isolated from strain KMS, as compared to the inferred amino acid composition of the TSST-1 structural gene.

^b Residues per mole values are based on a molecular weight of 22,000.

^c Residues per mole inferred from the DNA sequence of the TSST-1 structural gene. Blomster-Hautamaa and colleagues.

^d ND. Not determined.

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Comparison of the primary sequences of the staphylococcal enterotoxins and their relatives is shown in Figure 1. The complete primary amino acid sequences of the staphylococcal enterotoxins and related proteins are shown aligned, with the exception of the sequences of the exfoliating toxins, which are shown aligned, with the exception of the sequences of the exfoliating toxins, which are shown aligned with each other, but not with the remaining toxins. The exfoliating toxin sequences are shown here for completeness, and because these toxins have properties related to those of the others (see below). Toxins shown are as follows: SEA to SEE, *Staphylococcus aureus* enterotoxins A to E; SPE A and C, *Streptococcus pyogenes* toxins A and C; TSST1, *Staphylococcus aureus* toxic shock - associated toxin; ETA and ETB, *Staphylococcus aureus* exfoliating toxins A and B. Data are from (9-17). Residues that are identical or that have changed to an amino acid with similar properties among at least two of the following: SEA, SEE, and SED, are highlighted in pink. Residues that are identical or that have changed to an amino acid with similar properties among at least two of the following: SEB, SEC1, and SED and at least two of SEB, SEC1, and SEC2, are highlighted in yellow. Single letter abbreviations for the amino acid residues are: A, Ala; C, Cys; D, Asp; E, Glu; F, Phe; G, Gly; H, His; I, Ile; K, Lys; L, Leu; M, Met; N, Asn; P, Pro; Q, Gln; R, Arg; S, Ser; T, Thr; V, Val; W, Trp; and Y, Tyr.

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There is evidence that indicates varying degrees of immunological relatedness between certain enterotoxins. Bergdoll, M.S., Borja, C.R., Robbins, R., Weiss, K.F., Infect. Immun., 4, 593, 1971; Bergdoll, M.S., Enterotoxins. In: Staphylococci and Staphylococci Infections ed. C.S.F. Easmon, C. Adlam 1, pp. 559-598, 1983, Landon, Academic; Freer, J.H., Arbutnot, J.P., Pharm. Ther., 19, 55, 1983. A considerable degree of cross reactivity exists for antisera raised against one enterotoxin and other enterotoxins. It has been considered that the enterotoxins may contain major cross reactive antigenic sites, while each individual enterotoxin possesses minor specific antigenic regions. Common precipitating antibodies were formed between SEA and SED. In addition, enterotoxins B and C can react immunologically with antisera against either toxin type. Immunologic cross reactivity between Streptococcal pyrogenic exotoxin A and Staphylococcal enterotoxins B and C, has been shown. These results suggest a conserved domain present in the three exotoxins. SEA, SEB, SEC, SED, TSST-1 and the pyrogenic exotoxins have also been shown to share considerable DNA and amino acid homology. The enterotoxins, the pyrogenic exotoxins and TSST-1 therefore appear to be evolutionarily related and all belong to a common generic group of proteins.

It should be noted that the two Streptococcal toxins SPEA and C are about as similar to each of the Staphylococcal groups as they are to each other. Exfoliative toxins are of similar size to SEB and SEA with similar modes of action. They share several points of sequence similarity to the Staphylococcal enterotoxins. Overall there are several stretches at which similarities are apparent throughout the total group comprised of Staphylococcal enterotoxins, Streptococcal pyrogenic exotoxins and Staphylococcal exfoliative toxins. The

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longest of these, located two-thirds of the way through the proteins, is similar to sequences found at the C-terminal end of the human and mouse invariant chain.

Invariant chain is a polypeptide associated with nascent MHC class II molecules. Class II molecules bind peptides and present them to T cells during immune responses. Indeed, many toxins bind to class II molecules. The shared sequences may indicate some or all of the invariant chain and toxin binding sites on class II molecules.

The known structural homology between the enterotoxins and Streptococcal pyrogenic exotoxin is further supported by the identity of clinical responses. It is known that this exotoxin induces hypotension, fever, chills and septic shock in man. It is hypothesized that this compound activates cytokines, such as interleukin 1, interleukin 2, tumor necrosis factor and interferon, and procoagulant activity which are the prime mediators of the clinical symptomatology. It is hypothesized that many other bacterial products are capable of inducing similar in vivo activity. Among potential tumoricidal agents which are likely candidates based upon structural homology or identity of clinical symptomatology are gram positive bacterial products, cell wall bacterial constituents such as peptidoglycans and various gram negative bacterial components to include meningococcal, pseudomonas and E. Coli products. While presently undemonstrated in animal systems, it is believed that these agents are likely to possess similar tumoricidal utility as those claimed here for the enterotoxins.

The recognition that the biologically active regions of the enterotoxins and SPEA were substantially structurally homologous enables one to predict synthetic polypeptide compounds which will exhibit similar tumoricidal effects.

Figure 2 illustrates the amino acid sequence homology of

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mature SPEA and Staphylococcus aureus enterotoxin B. The top sequence is the SPEA-derived amino acid sequence. The amino acid sequence of enterotoxin B is on the bottom.

Sequences are numbered from the amino acid terminus, with amino acids represented by standard one character designations. (See Tables 5 and 6 below.) Identities are indicated by : and gaps in the sequences introduced by the alignment algorithm are represented by dashed lines. See Johnson, L.P., L'Italien, J.J., and Schlievert, P.M., "Streptococcal pyrogenic exotoxin type A (scarlet fever toxins) is related to staphylococcus aureus enterotoxin B," Mol. Gen. Genet. (1986) 203: 354-356.

One common methodology for evaluating sequence

homology, and more importantly statistically significant similarities, is to use a Monte Carlo analysis using an algorithm written by Lipman and Pearson to obtain a Z value. According to this analysis, a Z value greater than 6 indicates probable significance, and a Z value greater than 10 is considered to be statistically significant.

Pearson, W.R., Lipman, D.J., "Improved tools for biological sequence comparison," Proc Natl Acad Sci U S A, April 1988, 85 (8) pages 2444-8; Lipman, D.J., Pearson, W.R., "Rapid and sensitive protein similarity searches," Science, March 22, 1985, 227 (4693) pages 1435-41.

In the present invention, synthetic polypeptides useful in tumoricidal therapy are characterized by substantial structural homology to enterotoxin A, enterotoxin B and streptococcal pyrogenic exotoxins with statistically significant sequence homology and similarity (Z value of Lipman and Pearson algorithm in Monte Carlo analysis exceeding 6) to include alignment of cysteine residues and similar hydropathy profiles.

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TABLE 5

Amino Acid	One-letter Symbol
Alanine	A
Arginine	R
Asparagine	N
Aspartic acid	D
Cysteine	C
Glutamine	Q
Glutamic acid	E
Glycine	G
Histidine	H
Isoleucine	I
Leucine	L
Lysine	K
Methionine	M
Phenylalanine	F
Proline	P
Serine	S
Threonine	T
Tryptophan	W
Tyrosine	Y
Valine	V

One-letter
Symbol

5

10

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20

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TABLE 6

10	20	30	40	50
STRPKPSQORSNLVKTPIYIPFRTVL-----VTHENKVSVDQLLSHDLIYNVS--				
ESQDPKPKDELKSS--K-PTGLKENKV-LYNDHVSAINVKSINFP--DLIYLYSIK				
10	20	30	40	50
60	70	80	90	
-----GPNVDKLTKELNQEHATLPKDKNVDIYGYEYHLCYLC-----ENAEARSAC				
DTKLG-NYDNRVREFNKNKDLADKYDVDFGANYIQ-CYFSKTKNTNIDSHENTERTC				
60	70	80	90	100
100	110	120	130	140
LYGGVTHNEGHNLEIPKX---IVKVSIDGISOISLSPDIEQINGNCSRIS-YTVRYLTI				
HYCGVTEHCNQLD---KYRSITVRVFEDCKNLLSPDVQTHKKVTAEQLDYLFTHLYV				
120	130	140	150	160
160	170	180	190	200
DNKQLYTNGSPSYETGYIKIPKNKESFMPDFPEPE--PTQSKYLYMIYKDNETLDSNTS				
KMKLYEPNNSPYETGYIKIPKIE-NEHSFYDMHPAPGNKPDQSKYLYMHNKDKMVDSDV				
170	180	190	200	210
210	220			
QIEVYLTXX				
KIEVYLTXX				
230				

The enterotoxins are presumed to function by affecting emetic receptors in the abdominal viscera which stimulate the emetic and diarrheal response.

These toxins also stimulate T lymphocyte mitogenicity, procoagulant, chemotactic activity, as well as cysteinyl leukotriene, lymphokine, serine protease and thromboglobulin production. Cytokines known to be induced by enterotoxins induce interferone, tumor necrosis factor, interleukins one and two. They suppress immune responses, augment natural killer cell cytotoxicity, enhance gram-negative endotoxin lethality and induce fever and

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hypotension. These additional properties are shared with the pyrogenic exotoxins of both *Staphylococcus aureus* and *Streptococcus pyogenes* and TSST-1. Synthetic polypeptides would also be expected to demonstrate similar responses.

The *Staphylococcal* enterotoxins A, B, C, D, E, toxic shock toxin (TSST-1), a product of mycoplasma arthritidis and Mls antigens provoke dramatic T cell responses. *Staphylococcal* enterotoxins are the most powerful T cell mitogens known eliciting strong polyclonal proliferation at concentrations 10^3 lower than such conventional T cell mitogens as phytohemagglutinin. SEA is the most potent T cell mitogen, stimulating DNA synthesis at concentrations of 10^{-13} to 10^{-16} M in the human system. All stimulate a large proportion of both murine and human CD4+ and CD8+ T cells. Activity of these mitogens is tightly restricted by the major histocompatibility complex (MHC) class II antigens. It is proposed that the *staphylococcal* enterotoxins, streptococcal pyrogenic exotoxins, exfoliative toxins and a product of mycoplasma arthritidis bind directly to the T cell receptor and to class II MHC. These two structures are brought into contact, thus stimulating T cell activation via the β region of the T cell receptor mimicking strong alloreactive response.

Many toxins have binding affinities for MHC class II molecules which are involved in stimulating T cells. For example, SEA has a K_d for human class II of about 3.2×10^{-7} M, SEB of 10^{-6} M and TSST-1 of 10^{-7} M. SEA and SEB probably bind to the same site on class II because they cross compete for binding. Exfoliative toxins bind only weakly or not at all to class II. SEB and TSST-1 have different binding sites on class II molecules.

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The structure of class II consists of two immunoglobulin-like domains located close to the cell membrane which supports a structure constructed from the NH_2 terminal regions of both polypeptides of the protein and comprise an extended β -pleated sheet supporting two alpha helices separated by a cleft. Peptides derived from foreign materials or from proteolysis of self proteins normally lie in this groove. It is this complex of MHC and peptide that stimulates T cells bearing alpha and beta receptors. Bacterial toxins do not normally bind to MHC molecules by occupying this groove and therefore do not behave like conventional peptide-MHC binding antigens. Toxins bind to three different class II proteins, namely DR, DP, DQ (or murine I-A, I-E). SEB and TSST-1 bind to DR and DQ alleles but not to DP. Toxin-class II complexes stimulate T cells. Most toxins bind preferentially to DR class II proteins, less well to DQ and not at all to DP.

Different DR alleles have different affinities for a few of the toxins most notably SEE. In the mouse, complexes of toxins plus I-E (murine DR equivalent) stimulate T cells more efficiently than complexes of toxins with I-A (murine DQ analog). There is also evidence for weak haplotype specificity, e.g., toxins bound to I-A^k stimulate T cells less well than toxins bound to I-A^d or I-A^b. *Staphylococcus aureus* toxins bind more efficiently to human class II proteins than to mouse. A likely location for toxin binding to MHC may be at the sides of class II where 2 wings, the ends of the β -pleated strands, extend to either side of the proteins.

A hypothetical structure for the complex of class II MHC T cell receptor and *Staphylococcal* enterotoxins and MHC protein is given in Figure 3.

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The Figure shows a class II MHC protein, diagrammed according to Bjorkman and co-workers and Brown and co-workers, in contact with a T cell receptor and a staphylococcal enterotoxin or Mls product. Ag is the probable site of binding of a conventional antigenic peptide.

Toxins stimulate T cells through V_{β} binding.

T cell receptors for antigenic peptides bound to MHC proteins are made up of 5 clonally variable components V_{α} , J_{α} , V_{β} , D_{β} , and J_{β} . Recognition of most conventional antigenic peptides bound to MHC proteins involve contributions from all the variable components of the T cell receptor.

In contrast, the toxins stimulate T cells almost exclusively via the V_{β} region of the T cell receptor. See Table 7 for binding of toxins to T cells bearing various V_{β} receptors.

TABLE 7

TOXIN	V_{β} SPECIFICITY	
	HUMAN	MOUSE
SEA	?	1, 3, 10, 11, 17
SEE	5.1, 6.1-3, 8, 18	11, 15, 17
SED	5, 12, ?	3, 7, 8.1-3, 11, 17
SEC1	3, 12, 14, 15, 17, 20	3, 7, 8.1-3, 17
SEC2	12, ?	3, 8.2, 8.3, 11, 17
SEC3	12, 13.1, 13.2, 14, 15, 17, 20	3, 8.2, 10, 17
TsST1	5, 12, ?	3, 7, 8.1, 8.2
ExFT	2	3, 15, 17
MAH	2	3, 10, 11, 15, 17
	?	6, 8.1, 8.2, 8.3

This property of selective stimulation of V_{β} is reminiscent of the endogenous superantigens called Mls antigens in the mouse. The pattern of V_{β} specificity of the different toxins corresponds

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specificity of the different toxins corresponds loosely with their groupings by sequence similarity. SEA, SED and SEE all stimulate murine T cells bearing $V_{\beta}11$ and SEE and SED both stimulate human T cells bearing $V_{\beta}5$. SEB and SECs stimulate mouse T cells bearing members of the $V_{\beta}8$ family and human T cells positive for $V_{\beta}12$. The exceptions are as follows: SED stimulates T cells bearing the $V_{\beta}8$ unlike SEA and SEE. Exfoliating toxin and TSST-1 which are not related by sequence have similar specificities for V_{β} both in mouse and humans.

Bacterial toxins and other superantigens do not bind to T cell receptors at those regions involved in binding to conventional antigenic peptides plus MHC. The superantigens engage V_{β} on an exposed face of V_{β} or a region predicted to be a β -pleated sheet and exposed on the side of the T cell receptor. This model predicts that toxins act as clamps engaging the sides of class II and V_{β} bringing into close proximity the surfaces of the T cell receptor and MHC that would contact each other during T cell recognition of conventional antigens bound in the groove of MHC. Proper confirmation must await x ray crystallographic resolution of the complex.

Neither class II nor toxins separately have affinities for the T cell receptors in question, but the combination of toxins and class II proteins do. Only if the complex peptide-MHC ligand has formed can it functionally engage the T cell receptor. The T cell activation via the $V_{\beta}B$ region of the T cell mimics strong alloreactive responses. This interaction occurs irrespective of whether the V_{β} is expressed on CD4+ or CD8+ T cells. This behavior is consistent with the known resistance of

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Staphylococcal enterotoxins to proteolysis even in acidified conditions.

Mice Express Endogenous Equivalent of the Enterotoxins.

5 T cells from some mice responded well to spleen cells from some other animals even though both responder and stimulator were identical at the MHC. The antigens are called minor lymphocyte stimulating antigens (Mls). There are many Mls-like products produced by mice controlled by non-linked loci. Mls products stimulate T cells bearing V β S. Mls-1^a in combination with mouse class II molecules stimulate nearly all T cells bearing mouse V β 6, 7, 8.1 and 9. A list of the Mls-like products and the V β S they engage is given in Table 8. Mls products have not yet been found in humans.

TABLE 8

Mls-like products identified in mouse.

LOCUS	V β specificity	MHC association
Mls-1 ^a	6, 7, 8.1, 9	Class II (except q)
Mls-2 ^a	3	Class II (except q)
Mls-3 ^a	3	Class II (except q)
?	5	I-E
?	7	I-E
?	11	I-E
?	17	I-E

A striking resemblance exists between T cell responses to Staphylococcal enterotoxins and T cell responses to the Mls locus. The Mls locus located on chromosome 1 and other similar genes on other (unknown) chromosomes have profound effects on T cells. Polymorphism at these loci elicits a strong

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primary mixed lymphocyte response between MHC identical and Mls disparate spleen cells in mice.

Mls products stimulate T cells bearing particular V β S almost regardless of the rest of the structure of the receptor on the T cell. This activity depends on the simultaneous expression by the presenting cell of class II proteins. Some class II products, most notably I-E molecules, present Mls products and bacterial toxins better than others. Mls appear to engage V β S at the same site on the exposed face of the polypeptide as toxins.

The similarities between properties of bacterial toxins and mouse Mls products might lead one to suggest a structural similarity. Mls products associate with class II and stimulate T cells via V β much like superantigens but the structure of Mls is unknown.

There are consequences for mice expressing Mls products. They cause deletion in the thymus for all prospective T cells bearing V β S with which they interact. Mice expressing Mls-1^a contain very few T cells bearing V β 6, 7, 8.2 or 9 and hence are deprived of 20% of their total potential T cell repertoire. Despite this they do not seem to be susceptible to disease.

Both Mls and enterotoxins show the following characteristics in common:

1. Both activate a high frequency of normal T cells exceeding that of conventional protein antigens.
2. Responding T cells are CD4⁺.
3. T cells of many specificities respond.
4. Both elicit responses of T cells expressing receptors having particular V β gene products.

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5. There is no MHC restriction of responding T cells.
6. Both require presentation by class II MHC.
7. IE and IA molecules on antigen presenting cells are required for immunologic effects.
8. Ontogenetic deletion of V_{β} or $CD4^{+}8^{+}$ cells is induced by both molecules.

These similarities are summarized in Table 9.

TABLE 9

Similarities between the T cell responses to Mls and SE and differences with responses to protein Ag ^a		Mls-1 ^b		SE		Proteins	
Characteristic of the T Cell Response to:		Mls-1 ^b		SE		Proteins	
High frequency of responding cells		Yes (-1:5)	Yes (-1:5)	Yes (-1:5)	Yes (-1:10 ⁴)		
Responding T cells CD4 ⁺		Yes	Yes	Yes	Yes ^c		
T cell receptor involved in response		Yes	Yes	Yes	Yes		
T cells of many specificities respond		Yes	Yes	Yes	No ^c		
V_{β} restriction of responding T cells		Yes	Yes	Yes	Yes ^c		
MHC restriction of responding T cells		No	No	No	Yes ^d		
Incompetent class II MHC alleles		Yes	Yes	Yes	Yes ^d		
1-E more involved than 1-A		Yes	Yes	Yes	No		
Ontogenetic deletion of V_{β} on CD4 ⁺ 8 ⁺		Yes	Yes	Yes ^e	No		
Processing required		?	?	No	Yes		
Pulsing APC stimulatory		?	?	Yes	Yes		
Pulsing T cell stimulatory		?	?	Yes	No		
Protein identified		No	No	Yes	Yes		

^aData on T cell responses to Mls and SE derived from this paper and Janeway et al. A detailed description of the SE themselves is found in Bergdoll.

^bT cells expressing CD8 respond only to proteins degraded within cells; extrinsic proteins are presented by class II MHC to CD4⁺ T cells.

^cT cell responses to protein antigens require all elements of the TCR, whereas those to Mls and SE appear to require only use of certain V_{β} segments.

^dPresentation of proteins is much more restricted in use of allelic forms of class II MHC molecules than is "presentation" of SE or Mls.

^eFrom Yagi and Janeway.

The striking functional similarity of

Staphylococcal enterotoxins and Mls suggests that the Mls may represent a protein with homology to Staphylococcal enterotoxins. It has been proposed

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that the Mls like Staphylococcal enterotoxins directly binds the TCR-CD4 complex via its V_{β} domain and to class II MHC molecules assembling a complex that is highly stimulatory for T cells. Hence, both Mls and Staphylococcal enterotoxins are thought to ligate class II MHC to the TCR:CD4 complex in such a way as to stimulate a large percentage of T cells with restricted V_{β} usage.

While the animal studies described herein were carried out with Staphylococcal enterotoxins A and B, based upon the observed structural and reactive similarities, it would be expected that similar results would be obtained with the other Staphylococcal enterotoxins, SPEA, TSST-1, mycoplasma arthritidis, Mls antigens and the synthetic polypeptides described above. Additional biological properties common to this group include their mitogenic effects, interferon, interleukin and tumor necrosis factor induction activity. Furthermore, all are capable of inducing fever and shock when given intravenously to rabbits or monkeys, and most of these have been implicated as potential pathogenic agents in the toxic shock syndrome.

Production And Isolation Of

Enterotoxins A, B, C, D, E and F

General Methods

Isolation and purification procedures for

enterotoxins contain numerous common steps. On the whole, growth of enterotoxin producing Staphylococcus aureus strains is similar in all cases. The most widely used general medium for the culture of these organisms contains 3% M2-amine Type A, or MAX, 3% protein hydrolysate powder, 0.0005% thiamine and 0.001% niacin. Optimum yields of the enterotoxins

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are obtained under controlled fermentation, where pH, temperature and oxygen tension are controlled. Typically, growth at 37°C for 18 to 24 hours is sufficient for maximum toxin yields. The yield of enterotoxin B and C₁ and C₂ will be up to several hundred µgrams (toxin)/ml (media), while the yield of other toxins will be only a few µgrams/ml.

All enterotoxins are secreted products.

Generally, they are produced during the logarithmic and stationary stages of cell growth. After growth, the producing cells are removed from the medium by centrifugation, and the toxin-containing supernatant is saved. If a large fermentation has been carried out, then the cells and supernatant can be quickly separated using a continuous flow centrifuge. To concentrate the toxins from the media, various methods, e.g., polyethylene glycol precipitation, or dialysis tubing precipitation, or hollow fiber concentration using membranes with selective molecular weight cutoffs can be used. To assess the purity of the isolated enterotoxin product, specific antisera to each of the toxins are used in appropriate quantitative immunoassays, e.g., radioimmunoassays or enzyme labelled immunoassays, hemagglutination, or precipitin reactions.

Enterotoxin Purification By Type

Enterotoxin B

The strain of *Staphylococcus aureus*, that is used for the production of SEB (*Staphylococcal enterotoxin B*) is e.g., S6 or 10-275. (Source: Dr. John Iandolo, Kansas State University, Manhattan, Kansas.) The medium containing the toxin is diluted twice with water adjusted to a pH of 6.4, and AmberLite CG-50 (200 mesh) cation ion-exchange resin is added to the

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toxin mixture. The toxin is eluted, dialyzed, then reappplied to the CG-50 column again. The eluted toxin is dialyzed, then applied to a column of carboxymethyl cellulose or CM-Sephadex. Unbound proteins are eluted with 0.03 and 0.04 molar sodium phosphate buffer. At this point, the toxin is essentially homogeneous. Using chromatofocusing techniques, the SEB may be further subdivided into several isoelectric species using polybuffer 96.

Enterotoxin A (SEA)

High SEA producers, e.g., *Staphylococcus aureus* 13M-2909 (Source: Dr. John Iandolo, Kansas State University, Manhattan, Kansas) are grown in the general medium that is made 0.2% in glucose. Initially, AmberLite CG-50 is used for batch isolation. After incubation, the toxin is eluted and dialyzed. The toxin is then loaded onto a CM-cellulose column and eluted with a linear gradient. The combined fractions are then loaded onto a hydroxylapatite column and eluted using a linear gradient. The fractions are lyophilized and chromatographed on a Sephadex-G-75 column. The toxins obtained from this procedure are greater than 99% pure, with a yield of approximately 20%.

Enterotoxin C₁ (SEC₁)

Culture supernatant from *Staphylococcus aureus* 137 (Source: Dr. Marcia Betley, University of Wisconsin, Madison, Wisconsin) is concentrated, dialyzed and lyophilized. The toxin product is then applied to a carboxymethyl cellulose column and eluted with a stepwise gradient. The toxin peak consists of a sharp peak with a trailing edge. The eluted toxin is concentrated and applied to Sephadex-

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G-75. The toxin elutes as a single peak. The toxin is then concentrated and run twice through a column of Sephadex-G-50. The eluate is dialyzed against water and lyophilized.

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Enterotoxin C₁ (SEC₁)

Culture supernatant from *Staphylococcus aureus* 361 (Source: Dr. Marcia Betley, University of Wisconsin, Madison, Wisconsin) is concentrated as for SEC₁ and dialyzed. The toxin is then applied to a carboxymethyl cellulose column. SEC₂ is eluted, lyophilized and resuspended in distilled water. The toxin is reappplied to a column of carboxymethyl cellulose and eluted with a linear gradient. The partially purified toxin is concentrated and applied to a Sephadex-G-75 column. The eluted toxin is concentrated and finally reappplied to a Sephadex-G-50 column. Recovery is about 40%, with purity exceeding 99%.

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Enterotoxin D (SED)

Staphylococcus aureus 1151M (Source: Dr. John Iandolo, Kansas State University, Manhattan, Kansas) is used for the production of enterotoxin B. The medium is similar to that used for SEA and SEB. After growth and removal of the cells, the pH of the supernatant is adjusted to 5.6 and applied to an AmberLite-CG-50 resin. The mixture is stirred for one hour, and the toxin is eluted and concentrated using 20% (W/V) polyethylene glycol, 20M. The concentrated toxin is dialyzed and applied to a carboxymethyl cellulose column. The toxin is eluted in a linear gradient and then rechromatographed on carboxymethyl cellulose. The toxin solution is

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concentrated and chromatographed on Sephadex-G-75. This step is repeated once.

Enterotoxin E (SEE)

Staphylococcus aureus strain FRI-236 (Source: Dr. John Iandolo, Kansas State University, Manhattan, Kansas) culture supernatant is concentrated and dialyzed. The toxin is then absorbed to a carboxymethyl cellulose column. The toxin is eluted in a stepwise fashion and concentrated. It is then chromatographed twice on Sephadex-G-75. To obtain highly purified SEE, it is necessary to chromatograph the toxin once more on G-75 in the presence of 6 molar urea.

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Enterotoxin F or Toxic Shock Syndrome Toxin-1 (TSST-1), TSST-1a and TSST-1b

Staphylococcus strain MN8 (Source: Dr. Patrick Schlievert, University of Minnesota, Minneapolis, Minnesota) is cultured overnight in dialyzable beef heart medium and precipitated from culture fluid by adding 4 volumes of absolute ethanol and storing for at least 2 days. The precipitate is collected by centrifugation and the pellet is suspended in water, recentrifuged and dialyzed to remove salts. The preparation is then electrofocused in a pH gradient of 3-10 using commercial ampholytes with the LKB Multiphor apparatus. The visible band containing TSST-1 is harvested and refocused in a pH 6-8 gradient yielding purified TSST-1.

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TSST-1a and 1b are isolated by one additional electrofocusing step. After focusing TSST-1 on the pH 6-8 gradient, approximately one-half of the Sephadex gel is removed from the anode end. The gel remaining on the cathode end, containing the TSST-1 5153

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band is repoured after the addition of two more grams of Sephadex gel and then refocused overnight using the remaining pH gradient. After electrofocusing in a pH 6-8 or 6.5-7.5 gradient, protein bands are located by the zymogen print method. Discrete bands are scraped off the plate and eluted with pyrogen free water from the Sephadex gel. Strain MN8 yields approximately 2 mg of each toxin per liter of culture fluid. For *Staphylococcus aureus* strains other than MN8, 200 µg of each toxin is obtained per liter of culture fluid. TSST-1a and 1b are proteins which migrate as homogeneous bands in SDS gels to a molecular weight of 22,000 with isoelectric points of 7.08 and 7.22, respectively.

With the changing technology of protein purification, new methods have been employed for the purification of certain enterotoxins from *Staphylococcus aureus*. Some of these methods are given here.

20 Enterotoxins A and C₂

A 10 ml culture of *Staphylococcus aureus* 11N-165 (SEA), *Staphylococcus aureus* 361 (Source: Dr. John Iandolo, Kansas State University, Manhattan, Kansas) (SEC₂) is grown overnight at 37°C. The removal of enterotoxin from the supernatant is carried out using QAE-Sephadex. The toxin is then eluted batchwise from the ion exchanger and recovered by filtration on a sintered glass funnel. The eluates are concentrated by ultrafiltration. The toxin is then passed through a Sephadex-G-100 column. Two peaks absorbing at 280 nm are eluted, with the latter containing the enterotoxin. The eluted toxin is concentrated and rerun on Sephadex-G-100. The overall recovery is about 30% for SEC₂ and 40 to 50%

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for SEA. Both toxins appear homogeneous by sodium dodecylsulfate polyacrylamide gel electrophoresis.

Enterotoxins A, C₁, and D

This method utilizes fast protein liquid chromatography (FPLC) and high resolution chromatofocusing Mono P column. Enterotoxins in media are concentrated and passed over a Sephadex-C₄ 75 column. The toxin containing fractions are pooled. For C₁ and D, the supernatants are passed over an AmberLite-CG-50 column, as described for SED, and the active fractions pooled. All three toxins are then placed in buffer for chromatofocusing and then separated using the MONO P column FPLC system. Since all of the toxins have isoelectric points in the range of 7 to 9, the polybuffer PBE-96 is used for elution. The purity of SEA, SEC₁ and SED is estimated to be 98, 95 and 80%, respectively. SEA elutes as two peaks at pH 8.8 and 8.6. SEC₁ also elutes as two peaks at pH 8.3 and 7.9, and SED elutes as three peaks at pH 8.6, 8.3 and 8.0.

Enterotoxins may also be produced in mutant strains of *Staphylococcus aureus* by expression of an enterotoxin producing gene in another bacteria or cell. Genetic material which appears to be in the chromosomal plasmid, or phage portion of the bacteria may be used for gene insertion procedures. Complete molecules or fragments with amino acid sequence homology to the parent enterotoxin may be produced with this technology. (Reviewed in Iandolo, J.J., Annu. Rev. Microbiol., 43, 375, 1989.) Moreover, mutagenic agents such as N-Nitroso compounds are capable of augmenting significantly the production of enterotoxins by some strains of *Staphylococcus*.

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Alpha Toxin

Staphylococcus aureus Wood 46 strain (Source: Dr. Sidney Harshman, Vanderbilt University, Nashville, Tennessee) is used and cultured in yeast extract dialysate medium. With the glass-pore bead method undialyzed yeast may be used together with casein, glucose, thiamin and nicotinic acid. The organism is incubated in medium for 24h at 37°C.

The culture supernatant is applied to a glass-pore bead column and adjusted to pH 6.8. A column of 5 x 20 cm is used for 3 liter batches and flow rates adjusted to 10-20 ml/min. The column is washed with 0.01M KH₂PO₄ pH 6.8 and then the alpha toxin is eluted with 1.0M KH₂PO₄ pH 7.5. Fractions are tested for the presence of alpha hemolysin by a rapid hemolytic assay using rabbit erythrocytes as substrate.

Streptococcal Pyrogenic Exotoxin (SPE)

(Erythrogenic toxin, scarlet fever toxin)

Streptococcus MV-5 strain (Source: ATCC 12351) has been the most widely used for toxin production and studies. A list of various strains to produce toxins A, B, and C has been published. The Kalbach S84 type 3 strain (Source: Dr. Joseph E. Alouf, Institute Pasteur-Unite Associee, Paris, France) is cultured and the supernatant is concentrated and stirred in calcium phosphate gel. Fraction S₁ is precipitated with 80% saturated ammonium sulfate.

The redissolved pellet is dialyzed and termed

Fraction S₂. This fraction is precipitated with 50-80% ammonium sulfate, resuspended in phosphate buffered saline (Fraction S₃), and gel filtered on a Bio-Gel P-100 column. The fraction corresponding to the volume eluted between 160 and 240 ml is collected and concentrated by ultrafiltration to about 20 ml in

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an Amicon PM10 Membrane (Fraction S₄). Fraction S₄ is then submitted to preparative isoelectric focusing (IEF) performed with a 100 ml column. The material which focuses at around pH 4.8 in a narrow peak is collected and dialyzed in an Amicon cell using PBS to eliminate ampholines and sucrose. The Fraction (S₅) constitutes purified pyrogenic exotoxin. Another electrophoretic form of SPE with a pI of 4.2 is often separated simultaneously with that of pI 4.8. Both forms show total cross reactivity against immune sera raised by rabbit immunization with fraction S₃.

The Fraction S₅ shows a single band by SDS-PAGE corresponding to a molecular weight of 28K.

Bioassays for determination of activity include erythematous skin test in rabbits or guinea pigs lymphocyte blast transformation. The toxin may also be detected by enzyme-linked immunosorbent assay (ELISA) or hemagglutination inhibition.

Experimental Animal Studies

1. Enterotoxins

Staphylococcus aureus strain FRI-722 was the production strain for SEA and strain I10-275 was the production strain for SEB. Both were grown in 3% enzyme hydrolyzed casein and 1% yeast extract at pH 6.6 at a temperature of 35-37°C for 16-20 hours. The culture medium is then filtered and incubated with Amberlite CG50 and the bound protein eluted with high ionic strength buffer (impure preparations). For further purification, this step is repeated and elution carried out with a low ionic strength to high ionic strength buffer gradient. The fraction containing SEB is concentrated, dialyzed and loaded onto a molecular sieve gel filtration column. The fraction containing SEB is concentrated and dialyzed

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against phosphate buffered saline pH 7.2, filter sterilized and frozen. Samples containing 5 and 10 mg/ml are tested in double diffusion

5 Immunoprecipitation assay using known standards of SEB and SEA with monospecific antisera. A single precipitation line was noted which showed a line of identity with known SEB and SEA, respectively. For enterotoxin B batches, polyacrylamide gel electrophoresis showed bands at 28,000 and a weaker band at 20,000 molecular weight. Purified enterotoxin A showed only a single band at 28,000 molecular weight.

10 High performance liquid chromatography (HPLC) profiles were obtained on a MAC PLUS controlling a Rainin Rabbit HPLC with a Hewlett Packard 1040 A Diode array detector and a Vydac Protein and Peptide C18 column. The profile for purified enterotoxin B was a sharp peak without significant shoulder. There was minimal trace contamination. A functional hemolytic assay for the presence of alpha hemolysin in the pure preparation was negative. Purified enterotoxin batches were negative for endotoxin in the limulus amoebocyte lysate assay. The sterility of the preparations was demonstrated by negative cultures in thioglycollate medium and soybean-casein digest. Protein determinations were carried out by a spectrophotometric method.

20 The functional activity of the toxins in vitro was tested using an established system of mitogenicity. Human peripheral blood lymphocytes 5×10^5 /well, were incubated in flat bottomed microtiter plates and stimulated with graded doses of exotoxins for various periods. Each well was pulsed with $1 \mu\text{Ci}$ (^3H) thymidine 18 hours before termination

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of the cultures which were harvested and processed for liquid scintillation counting.

2. Animals

5 New Zealand white female rabbits weighing from 2.5 to 5.0 kg, ages 2 to 4 months were used for studies employing purified enterotoxins. Rabbits of higher weight were used in the preliminary studies which are discussed in application Serial No. 07/416,530, filed on October 3, 1989. The animals were obtained from the Elkhorn Rabbitry, Watsonville, California.

3. Tumor

15 The tumor used for these studies was obtained from the Frederick Cancer Research Facility of the National Cancer Institute. It was stored frozen in the DCT tumor repository. The tumor call lettered G50014 was also known as the VX-2. Stewart, H.L., Snell, K.C., Dunham, L.J.: Transplantable and transmissible tumors of animals. In Atlas of Tumor Pathology. Washington, D.C, Armed Forces Institute of Pathol., pp. 38, 355, 1959. The tumor is a carcinoma indigenous to the New Zealand white rabbit. It was stored as a tissue fragment, and suspended in saline. The tumor was initially induced by Shope virus and derived from a transformed papilloma in a dutch belted rabbit. Kidd and Rous described the tumor in 1937. Histopathologically, the tumor consists of cords and sheets of epithelial cells (80%) and 20% hemorrhage and necrosis with no acini. The growth is primarily papillary. Numerous mitoses are evident. The cells are thin walled and very anaplastic. The tumor used was cryopreserved from October 20, 1985. It had a negative viral profile.

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4. Tumor inoculation

Tumor fragments for inoculation were obtained from VX-2 growing in rabbit thigh. Fragments were implanted intramuscularly into the right thigh of recipients. Donors were placed under general anesthesia with halothane (1.5%) and under sterile conditions, small fragments were excised and placed in Dulbecco's Modified Eagles Medium with glutamine (Gibco Life Technologies, Inc., Grand Island, NY 14072). The fragments were rinsed and then suspended in media until they were transferred into new hosts. Recipient rabbits had their right thigh shaved and scrubbed with alcohol and betadine. A small area was anesthetized with 1% lidocaine. With a scalpel, an incision was made through the skin into the muscle where a small pocket was created. With forceps, 4 to 5 tumor fragments were implanted into the muscle. The wound was closed with 1 or 2 nylon sutures. Tumors appeared at the implantation site within 4 weeks and therapy was started when the tumors were at least 1 to 2 centimeters in broad diameter.

5. Tumor measurements.

Tumors were measured by calipers by a certified veterinary oncologist before and at intervals after treatment. Complete remission was present when there was no evident tumor. Partial remission represented a reduction of tumor volume by greater than 50%. Less than partial remission was a 25-50% reduction in tumor volume.

6. Conditions of Administration

Enterotoxins A and B in lyophilized form were diluted in 0.9% saline and then filtered through a 0.45 micron Millipore filter. Aliquots were stored at -20° F. An aliquot was thawed once, used only for a single injection and then discarded. Enterotoxin preparations in appropriate dose were prepared in 1 ml of 0.9% saline and drawn up in a 1 ml syringe. This solution was administered via the central ear vein which was cannulated with a 25 gauge needle and attached infusion tubing (Butterfly, 25 x 3/4 with 12" tubing set, Abbott Hospital, N. Chicago, IL 50064). The tubing and needle were initially washed with saline using a 3 ml syringe and with the tubing filled with saline, the toxin infusion was begun using a 1 ml tuberculin syringe (Monoject tuberculin 1.0 cc, Division of Sherwood Medical, St. Louis 63103). Approximately 0.3 ml of toxin was administered per minute. The tubing and needle were washed with 6 ml of normal saline over an additional 3 minutes using a 3 ml syringe. Partially purified enterotoxin B in doses of 100-150 µg enterotoxin/kg body weight or 40-60 µg enterotoxin/kg body weight were injected. Purified enterotoxin B was used in doses of 13-40 µg enterotoxin/kg body weight and purified enterotoxin A was used in doses of 0.5-12 µg enterotoxin/kg body weight.

7. Enterotoxin Administration to Tumor Bearing Rabbits.

Preliminary studies in 20 rabbits using partially purified enterotoxin B as a single dose of 100-150 µg/kg or 40-60 µg/kg resulted in tumor regressions. With a dose of 40-60 µg/kg, six of twelve animals showed objective tumor regressions while a dose of 100-150 µg/kg resulted in objective tumor responses

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in three of nine rabbits treated. Results of these studies are given in prior application, Serial No. 07/466,577, filed on January 17, 1990. Toxicity of these preparations was thought to be due to contaminating elements in particular staphylococcal alpha hemolysin. Accordingly, the next phase of these studies was carried out with purified enterotoxin B.

a) Purified Enterotoxin B

Purified enterotoxin B in a mean dose of 26 $\mu\text{g/kg}$ was administered to seven animals on one, two or three occasions (Table 10). Five showed complete remissions while one additional rabbit demonstrated 96% regression. One showed tumor progression. Of the four animals receiving a mean dose of 13 $\mu\text{g/kg}$, one had a complete remission while three showed tumor progression. A single animal given a dose of 40 $\mu\text{g/kg}$ died within 12 hours of injection. Six of eight animals with major regressions showed enduring responses lasting 2 to 6 months without evident tumor recurrence (Table 10).

b) Purified Enterotoxin A

SEA in a dose of 0.5 $\mu\text{g/kg}$ was given to 5 rabbits on two or three occasions. One showed a complete remission and a second 93.5% remission while two others demonstrated tumor progression and one died acutely after the third injection. SEA in a dose range of 5-12 $\mu\text{g/kg}$ was administered to 7 animals. Two achieved complete remission while one experienced a 60% remission. Four others died acutely after the first injection.

c) Purified TSST-1

One rabbit was treated with 0.5 $\mu\text{g/kg}$ of TSST-1 and showed a complete remission over 40 days.

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d) Untreated Control Animals

Five animals were inoculated with the VX-2 carcinoma as given above but were not treated with enterotoxins. All five showed progressive tumor growth over 90 days observation. No spontaneous remissions of tumor were observed.

TABLE 10

Purified Enterotoxin B (Lot TTB-16)

Animal Number	Maximum Response	Time to Maximum Response (days)
---------------	------------------	---------------------------------

Mean Dose: 26 $\mu\text{g/kg}$

Or ²	complete remission	24
Wanda ¹	complete remission	20
Cindy ²	complete remission	30
Edna ³	complete remission	46
Magnolia ³	complete remission	75
Periwinkle ³	96% progression	68

Mean Dose: 11 $\mu\text{g/kg}$

KT ¹	complete remission	14
Dinky ²	progression	
Hazle ²	progression	
Gretta ¹	progression	

Mean Dose: 40 $\mu\text{g/kg}$

Bonnie ¹	NC	12 hours (acute death)
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1 One injection on day 0.

2 Two injections: One injection on day 0 and one injection on days 4, 5, 7 or 8.

3 Three injections: One injection on day 0, one injection on days 4 or 6, and one injection on days 11 or 13.

8. Histology

Microscopically, tumors showed extensive

hemorrhagic necrosis in samples obtained 12 to 72

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hours after the initial injection. Control untreated tumor showed focal areas of necrosis within the tumor, but no areas of hemorrhagic necrosis. Indeed, the areas of necrosis were far more extensive in the treated tumors with few if any areas of viable tumor. In the treated tumors, small blood vessels demonstrated hemostasis, and focal areas of inflammatory cell extravasation in the perivascular area. These changes were not seen in control untreated tumor specimens.

9. Toxicity

Acute toxic effects were observed in all animals after enterotoxin administration, to include temperature elevations of 2 to 5°F and anorexia. The animals became subdued. At the same time, the tumors became soft and mobile. There were signs of diarrhea in several animals. Toxicity of partially purified enterotoxin B is given in copending patent application Serial No. 07/416,530, filed October 3, 1989.

a) Purified Enterotoxins A and B

With purified enterotoxin B, the dose range of 5-26 µg/kg was associated with fever and anorexia, while the 40 µg/kg dose was lethal in one animal tested. Five of seven rabbits given enterotoxin A in doses of 5-12 µg/kg died within 72 hours of the first dose. However, when the dose was reduced to 0.5 µg/kg four of five animals survived with two showing tumor regressions and one dying after the third injection. The surviving animals showed temperature elevations of 2° to 5° F and anorexia for 1-3 days after injection. During acute inflammatory activity in the tumor, animals often lost weight.

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10. Multiple injections of enterotoxins induce antitumor effects

Tumor bearing rabbits were given two or three injections of SEB, SEA or TSST-1 and showed tumor regressions. It is known that enterotoxins induce production of various cytokines and that one such cytokine namely interferon will in turn upregulate the surface expression of IA molecules and Class II major histocompatibility antigens. Such additional upregulated antigen presenting cells, would be further capable of binding additional enterotoxins and presenting them to the T lymphocyte repertoire. Moreover, a synergy has been noted between various cytokines namely tumor necrosis factor, interferon and various mitogens for T lymphocyte activation. Therefore, we may speculate that in the presence of various cytokines induced by the first injection of enterotoxins, upregulated antigen presenting cells are primed to bind additional toxin given in the second or third injection producing substantially augmented T cell proliferative responses and associated anti-tumor effects.

It is conceivable that the enterotoxins might be employed together with various cytokines such as IL-2 *in vitro* to develop a highly enriched population of T lymphocytes that could subsequently be injected at various intervals to continuously augment the anti-tumor effect in tumor bearing hosts.

Finally, while the enterotoxins were given intravenously in the present experiments, it is quite conceivable that the toxins could be administered in adjuvant form bound to vehicles such as aluminum hydroxide, liposomes, water in oil emulsions, pluronic triblock polymers and saponin with similar anti-tumor effects.

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11. Attenuation of toxicity with Ibuprofen

The administration of Ibuprofen (20 mg/ml) given in doses of 0.25 to 0.5 ml subcutaneously when temperatures reached 105°F or greater resulted in reduction in fever by 2 to 5°F. Ibuprofen could be administered every 4 to 6 hours; however, in general, it did not need to be given more than once or twice per 24 hours. The use of this drug did not interfere with the observed tumor reduction or histologic hemorrhagic necrosis.

Ibuprofen may inhibit the prostaglandin mediated effects of the inflammatory cytokines including fever and anorexia but does not affect other antitumor immune and inflammatory responses.

Ibuprofen is only one of a large group of drugs known as non-steroidal anti-inflammatory agents (cyclooxygenase and prostaglandin synthesis inhibitors), which would also be useful to attenuate toxicity induced by the enterotoxins.

12. Genetic Aspects of Enterotoxin Production

Proceeding from the seminal work of Cohen & Boyer, U.S. Patent No. 4,237,224, rDNA technology has become useful to provide novel DNA sequences and produce large amounts of heterologous proteins in transformed cell cultures. In general, the joining of DNA from different organisms relies on the excision of DNA sequences using restriction endonucleases. These enzymes are used to cut donor DNA at very specific locations, resulting in gene fragments which contain the DNA sequences of interest. These DNA fragments usually contain short single-stranded tails at each end, termed "sticky-ends". These sticky-ended fragments can then

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be ligated to complementary fragments in expression vehicles which have been prepared, e.g., by digestion with the same restriction endonucleases. Having created an expression vector which contains the structural gene of interest in proper orientation with the control elements, one can use this vector to transform host cells and express the desired gene product with the cellular machinery available. Once expressed, the gene product is generally recovered by lysing the cell culture, if the product is expressed intracellularly, or recovering the product from the medium if it is secreted by the host cell.

Recombinant DNA technology has been used to express entirely heterologous gene products, termed direct expression, or the gene product of interest can be expressed as a fusion protein containing some parts of the amino acid sequence of a homologous protein. This fusion protein is generally processed post-translationally to recover the native gene product. Many of the techniques useful in this technology can be found in Maniatis, T., et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, New York (1982).

However, while the general methods are easy to summarize, the construction of an expression vector containing a desired structural gene is a difficult process and the successful expression of the desired gene product in significant amounts while retaining its biological activity is not readily predictable. Frequently gene products are not biologically active when expressed in yeast, bacteria or mammalian cell systems. In these cases, post-translational processing is required to produce biological activity.

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From physical and genetic analysis, the genes for SEA, SEB, SEC, and SEE occupy a chromosomal loci. The structural gene encoding SED in all strains examined is localized, to a large penicillinase-like plasmid.

The enterotoxin A gene has been cloned. SEA was expressed in the *E. coli* genetic background from a single 2.5 kbp Hind III chromosomal DNA fragment. When sequenced, the DNA was found to contain a single reading frame that generated a protein consistent with the partial sequences of SEA derived by chemical methods. Therefore, it is apparent that the site mapped contained the structural gene for SEA.

Betley, M.J., Mekalanos, J.J., *J. Bacteriol.* 170, 34, 1987; Huang, I.Y., Hughes, J.L., Bergdoll, M.S., Schantz, E.J., *J. Biol. Chem.*, 262, 7006, 1987; Betley, M., Lofdahl, S., Kreiswirth, B.N., Bergdoll, M.S., Novick, R.P. *Proc. Natl. Acad. Sci., USA*, 81, 5179, 1984.

The enterotoxin A gene was found to be at least 18 kilobases in length and was carried on a mobile element. Enterotoxin A production was linked to the presence of a bacteriophage which integrates into the bacterial chromosome. The enterotoxin A gene is located near the phage attachment. The enterotoxin A gene was mapped between the purine and isoleucine-valine markers in 24 *Staphylococcus aureus* strains. Conversion to the SEA producing phenotype was induced by lysogenization with a temperate phage purified from *Staphylococcus aureus* strain PS42D. Therefore, a bacteriophage vector was found to be responsible for the toxin phenotype in suitable recipients.

The enterotoxin B gene has been cloned and expressed in *E. coli*. The DNA of the gene derived from *E. coli* has been sequenced and matches the

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chemically derived sequence with only minor differences. Gaskill, M.E., Khan, S.A., *J. Biol. Chem.*, 263, 6276, 1988; Jones, C.L., Khan, S.A., *J. Bacteriol.*, 166, 29, 1986; Huang, I.Y., Bergdoll, M.S., *J. Biol. Chem.*, 245, 3518, 1970.

The SEC gene has been cloned from the chromosome of *Staphylococcus aureus* MN Don. The cloned toxin was expressed in *E. coli* with a molecular weight comparable to that of the toxin from *Staphylococcus aureus*. The toxin was biologically active as measured by pyrogenicity, enhancement of lethal endotoxic shock and mitogenicity with murine splenocytes. The DNA sequence of the enterotoxin C gene has been developed and a protein sequence derived that compares favorably with the complete chemical sequence reported earlier. Bohach, G.A., Schlievert, P.M., *Infect Immun.*, 55, 428, 1987; Bohach, G.A., Schlievert, P.M. *Mol. Gen. Genet.* 209, 15, 1987.

The enterotoxin D gene has been found to occur on a 27.6 kbp plasmid. The enterotoxin D gene has been cloned and expressed in *E. coli* and other *Staphylococcus aureus* strains. The enterotoxin D gene in *Staphylococcus aureus* is under control of the agar locus like most *Staphylococcus aureus* extracellular protein genes. The DNA sequence was determined encoding a mature protein with amino acid composition and reaction with antibody to SED confirming its identity to the biochemically purified toxin. Couch, J.L., Saltis, M.T., Betley, M.J., *J. Bacteriol.* 170, 2954, 1988.

The enterotoxin E gene has been cloned from *S. aureus* FRI918 and was expressed in *E. coli* encoding an extracellular protein of 26,425 daltons. Its identity to SEE was confirmed immunologically and by

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correspondence of N terminal and C terminal analysis. Kreiswirth, B.N., Lofdahl, S., Betley, M.J., O'Reilly, Schlievert, P.M. Nature, 305, 709, 1983.

TSST-1 gene was not associated with either bacteriophage or plasmid DNA. The gene was cloned on a 10.6 kbp fragment of chromosomal DNA and subsequently on an approximately 1 kbp subclone of the larger fragment. The TSST-1 gene was expressed in *E. Coli*, and TSST-1 was secreted into the periplasm. The genetic element coding for TSST-1 was found to occupy two loci on the *Staphylococcus* genome. The loci are indicated by the notation H1 555; one is at the junction of regions 1 and 2 and is indistinguishable from att012 and closely linked to *tyrB*. The second is within the *trp* operon at the junction of regions 17 and 18. H1555 encodes the *tst* gene and is a heterologous insertion element that provisionally exhibits some of the characteristics of a transposon. Strains that are *Trp*⁻ contain H1555 at regions 17 and 18 (linked to *Trp*), while strains that are *Trp*⁺ contain H1555 elsewhere linked to *TryB*. However, the *Trp*⁻ phenotype is not due to insertional inactivation by the unusual element. The sequence and analysis of the *tst* gene has been described. It codes for a mature protein (TSST-1) of 197 amino acids and a molecular weight of 22,049. Cooney, J., Mulvey, M., Arbuthnott, J.P., Foster, T.J., J. Gen. Microbiol., 134, 2179, 1988.

Streptococcal pyrogenic exotoxin (SPEA) is clearly related to the enterotoxins. It has a cysteine loop of 9 amino acids similar to that of SEA and is also encoded by a converting phage. SPEA shows greater amino acid sequence similarity with SEB than SEA. Immunologic studies show that the proteins and antisera to either enterotoxin are cross

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reactive. Therefore, genes for all of the enterotoxins have been isolated and transfected into other bacteria to obtain selective production. These genes may be used as sources of accelerated production of these toxins in high producing bacteria employing transfection techniques familiar to one skilled in the art. Iandolo, J.J., Annu. Rev. Microbiol., 43, 375, 1989.

High producing strains of *Staphylococcus* for selective enterotoxin production have been identified and are available as described in enterotoxin purification section above. Moreover, exposure to mutagenic agents such as N-methyl-N'-nitro-N-nitrosoguanidine of enterotoxin producing *Staphylococcus aureus* has resulted in a 20 fold increase in enterotoxin production over the amounts produced by the parent *Staphylococcus aureus* strain. Freedman, M.A., Howard, M.B., J. Bacteriol, 106, 289, 1971.

13. Transfections of Tumor Cells With Enterotoxin Genes

The genes for the enterotoxins and streptococcal pyrogenic exotoxins have been cloned. With their known mimicry of the *Mis* locus and their affinity for T cell beta receptors, it would be logical to assume that transfection of the enterotoxin gene into tumor cells bearing appropriate HLA-DQ or DR or DP would result in production of a tumor cell bearing the minor lymphocyte stimulating locus capable of ligating MHC class II molecules with T lymphocytes, therefore stimulating potent T cell proliferation and associated antitumor immunity.

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14. Bacterial Products Related to
Staphylococcal Enterotoxins With Similar
Biological Effects

Streptococcal pyrogenic exotoxin (SPE) is
produced by many strains of group A streptococci.
Three antigenically distinct types (A, B, C) have
been described. It is now known that Streptococcal
pyrogenic exotoxin or scarlet fever toxin is related
to Staphylococcus aureus enterotoxin B. The amino
acid sequence of SPE has significant homology with
Staphylococcus aureus enterotoxin B but not with
other proteins in the Dayhoff library. Figure 2
shows the alignment of amino acid sequences of mature
SPEA and Staphylococcus aureus enterotoxin B, as
reported in Johnson, L.P., L'Italien, J.J. and
Schievert, P.M. "Streptococcal pyrogenic exotoxin
type A (Scarlet fever toxin) is related to
Staphylococcus aureus enterotoxin B," Mol. Gen. Genet.
(1986) 203:354-356.

The biological properties of SPE are shared with
some Staphylococcal enterotoxins such as lymphocyte
mitogenicity, fever induction and enhanced
susceptibility to endotoxin shock when given
intravenously. SPE activates murine T cells mainly
V β 8.2 in physical association with MHC class II
molecules expressed on accessory cells. SPE causes
deregulation of the immune response *in vitro*
resulting in delayed (12-16 days) acceleration of
humoral and cellular immune activity. This may
account for the sustained anti-tumor responses noted
with the use of its structural analog, namely
enterotoxin B, when administered to rabbits with the
VX2 carcinoma as demonstrated herein. Moreover, SPE
has now been shown to induce a toxic shock like
syndrome identical to that associated with various
enterotoxins. Given the biological and structural

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relatedness of these proteins, it would be
anticipated that SPE and any other protein, bacterial
or otherwise, with homology to enterotoxins would
produce tumoricidal effects identical to those of
enterotoxins.

15. Staphylococcal Enterotoxin Protein
Fragments With Biologic Activity

Studies of amino acid homology of Streptococcal
pyrogenic exotoxin and enterotoxin B have suggested
that there may be biologically active fragments
present within the whole molecule. Indeed, cyanogen
bromide generated toxin fragments of TSST-1 have been
shown to be responsible for T lymphocyte mitogenicity
and suppression of immunoglobulin synthesis. These
functions could be selectively blocked by monoclonal
antibodies directed to the respective fragments.
Amino acid analysis of the toxins show that they
contain similar domains that may give rise to
mitogenic and emetic properties in susceptible cells.
A peptide fragment in SEC was shown by Spero and
Morlock to contain the active sites for emesis and
diarrhea. The mitogenic region resided in the C
terminal tryptic fragment of SEC.

An immune functional site on Staphylococcal
enterotoxin A has been identified corresponding to
residues 1-27 of SEA which is responsible for
stimulation of T cell proliferation and induction of
interferon- γ . This SEA (1-27) sequence corresponds
to N-Ser-Gly-Lys-Ser-Glu-Glu-Ile-Asn-Gly-Lys-Asp-
Leu-Arg Lys-Lys-Ser-Glu-Leu-Gln-Gly-Thr-Ala-Leu-Gly-
Asn-Leu-Ly and blocks SEA induced T cell
proliferation and production of interferon γ which
was not seen with SEA (28-48) peptide. Thus, a
functional site on SEA responsible for modulation of

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T cell function involves the N-terminal 27 amino acids. These molecules may interact at either the level of TCR or the binding of SEA to class II MHC antigens.

For TSST-1, mitogenic activity was shown to be located on a 14,000 dalton cyanogen bromide generated toxin fragment. Other studies using proteolytic digestion of the TSST-1 with papain demonstrated mitogenic activity in 12,000 dalton fragment occupying 2/3 of TSST-1 molecule toward COOH terminal end of holotoxin. On the other hand, non-specific mitogenicity of rabbit lymphocytes demonstrated by enterotoxins A, B, and C₁ was associated with the NH₂ terminal ends of the molecules.

The emetic reaction and a related immediate-type skin reaction to SEB appears to be mediated by histamine and cysteinyl leukotrienes liberated from mast cells. Enterotoxins probably act on intramucosal or intradermal ganglion cells and the effect on mast cells is indirectly mediated by neuropeptides. Carboxymethylation of histidine residues of SEB caused a complete loss of emetic and skin sensitizing activity without changing the immunological specificity, e.g., T cell stimulating activity. An anti-idiotypic monoclonal antibody against the combining site of an anti-SEB monoclonal antibody had no enterotoxic activity but can inhibit the enterotoxic activity, e.g., emetic response and diarrhea of a 10,000 molar excess of SEB. Anti-idiotypic antibody also inhibited immediate-type skin reactions as well. The anti-idiotypic antibody and carboxymethylated enterotoxins may be useful tools to protect against the enterotoxin induced intestinal toxicity.

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Therefore, it could be predicted that fragments of the whole enterotoxin molecule can produce biologically active effects and predictably reproduce the in vivo tumoricidal activity of the whole molecule while eliminating some of the toxic effects noted.

Moreover, it would be reasonable to assume that similar or increased tumoricidal effects could be accomplished with biologically active fragments or intact enterotoxins alone or attached to antigen presenting cells (class II MHC, HLA-DR) and incubated ex vivo with a random T cell population or one which may have been pre-enriched for the appropriate V_H receptor. The activated T cell population with bound enterotoxin might then be reinfused into the host. Similar tumoricidal effects would be anticipated with enterotoxins or biologically active fragments infused into a host who has had an "organoid" (an enriched T lymphocyte organ) implanted on a biocompatible matrix and placed in a site in the host such as the abdominal cavity, adjacent to the liver or subcutaneously.

Although the foregoing invention has been described in detail for purposes of clarity of understanding, certain modifications may be practiced within the scope of the appended claims. While the above findings apply to an experimental animal model, it should be recognized that the tumor used herein is an excellent model of human cancer. Therapeutic success in the canine model with PACC system (described in a series of patent applications, the latest of which is identified as Serial No. 331,095), the forerunner of the present invention, was transferred to humans in which objective tumor regressions were obtained in four of the first five

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consecutive patients treated. Thus, the data given herein for rabbits with carcinoma is expected to be predictive of success when the compositions are applied to humans with spontaneous tumors as well.

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CLAIMS

1. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins produced by secretion from Staphylococcus aureus during growth phase, isolated from growth media by precipitation or centrifugation, and purified by chromatographic techniques.
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2. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, obtained by isolation from high enterotoxin producing mutant strains of Staphylococcus aureus.
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3. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxin fragments, obtained by isolation from high enterotoxin producing mutant strains of Staphylococcus aureus.
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4. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, obtained by isolation from high enterotoxin producing bacteria capable of expressing Staphylococcus aureus enterotoxin genes.
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5. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxin fragments, obtained by isolation from
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high enterotoxin producing bacteria capable of expressing Staphylococcus aureus enterotoxin genes.

6. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of Staphylococcal enterotoxins, obtained by isolation from high enterotoxin producing cells capable of expressing Staphylococcus aureus enterotoxin genes.

7. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of Staphylococcal enterotoxin fragments, obtained by isolation from high enterotoxin producing cells capable of expressing Staphylococcus aureus enterotoxin genes.

8. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of Staphylococcal enterotoxins obtained by isolation from Staphylococcus aureus which has been treated with chemical mutagens to obtain high enterotoxin producing strains.

9. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of Staphylococcal enterotoxins fragments obtained by isolation from Staphylococcus aureus which has been treated with chemical mutagens to obtain high enterotoxin producing strains.

10. A method of treating cancer in a patient which comprises administering to the patient a

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tumorcidally effective amount of Staphylococcus enterotoxins which have been chemically derivatized to minimize or delete emetic and skin sensitizing toxicity while retaining T cell and cytokine stimulating activity.

11. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of staphylococcal enterotoxins utilizing single or multiple injections.

12. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of staphylococcal enterotoxins incorporated in an adjuvant selected from the group consisting of aluminum hydroxide, liposomes, water in oil emulsions, pluronic triblock polymers, and saponin.

13. A method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of staphylococcal enterotoxin bound to solid supports to stimulate and enrich a T lymphocyte population in vitro with antitumor activity that may be subsequently administered to a tumor bearing host.

14. The method of treating cancer in a patient which comprises administering to the patient a tumorcidally effective amount of staphylococcal enterotoxin used together and synergizing with cytokines such as interferon and tumor necrosis factor in vitro to induce augmented T lymphocyte proliferation and antitumor activity that may be subsequently administered to a tumor bearing host.

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15. The method of Claims 1, 2, 3, 4, 5, 6, 7, 8 or 9 in which the tumorically effective amount of Staphylococcal enterotoxin is between 0.5 µg (enterotoxin) / kg (body weight) and 150 µg (enterotoxin) / kg (body weight).

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16. The method of Claims 1, 2, 3, 4, 5, 6, 7, 8 or 9 further comprising administering a toxicity attenuating dosage of a non-steroidal, anti-inflammatory agent, and cyclooxygenase and prostaglandin synthesis inhibitor to said patient.

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17. A method of treating cancer in a patient which comprises administering to the patient a tumorically effective amount of Streptococcus pyrogenic exotoxin.

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18. The method of Claim 12 in which the tumorically effective amount of Streptococcus pyrogenic exotoxin is between 2 µg (exotoxin) / kg (body weight) and 150 µg (exotoxin) / kg (body weight).

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19. The method of Claims 12 or 13 further comprising administering a toxicity attenuating dosage of a non-steroidal, anti-inflammatory agent and prostaglandin synthesis inhibitor to said patient.

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20. The method of Claim 15 further comprising administering a toxicity attenuating dosage of non-steroidal, anti-inflammatory agent and cyclooxygenase and prostaglandin synthesis inhibitor to said patient.

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21. The method of Claim 16 further comprising administering a toxicity attenuating dosage of non-steroidal, anti-inflammatory agent and cyclooxygenase and prostaglandin synthesis inhibitor to said patient.

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22. A method of simultaneously activating cytokine mediators such as Interleukin 1, Interleukin 2, tumor necrosis factor and interferon, and procoagulant activity which comprises administering to the host a tumorically effective amount of streptococcus pyrogenic exotoxin.

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23. A method of simultaneously activating cytokine mediators such as Interleukin 1, Interleukin 2, tumor necrosis factor and interferon, and procoagulant activity which comprises administering to the host a tumorically effective amount of staphylococcal enterotoxins.

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24. A method of simultaneously augmenting natural killer cell cytotoxicity and activating cytokine mediators such as Interleukin 1, Interleukin 2, tumor necrosis factor, interferon and procoagulant systems and stimulating T-lymphocyte mitogenicity, serine protease and thromboglobulin production by administering a tumorically effective amount of staphylococcal enterotoxins.

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25. A method of treating cancer in a patient which comprises administering to the patient a tumorically effective amount of molecule(s) functionally and structurally similar to Staphylococcal enterotoxins which have the properties

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of superantigens, binding to both TCR-CD4 complex of T lymphocytes via the V_H domain and to class II MHC molecules (IE or IA bearing) on antigen presenting cells thus assembling a complex that is highly stimulatory for T cells.

26. A method of inducing a tumoricidal response with enterotoxins A, B, C, D, E and F and pyrogenic exotoxins and biologically active fragments of enterotoxins by attaching these to antigen presenting cells and incubating with random or enriched T cell populations and the reinfusing of the activated cells into the host.

27. A compound useful in tumoricidal therapy consisting of a synthetic polypeptide with substantial structural homology to Staphylococcal enterotoxins and streptococcal pyrogenic exotoxins with statistically significant sequence homology and similarity to include alignment of cysteine residues and similar hydrophathy profiles.

28. A method of treating cancer in a patient which comprises administering to the patient a tumoricidally effective amount of tumor cells which have been transfected with the enterotoxin gene resulting in a transformed cell with surface expression of the minor lymphocyte stimulating locus equivalent and consequent potent activation and proliferation of T lymphocytes, especially those with V_H receptors and anti-tumor responses.

AMENDED CLAIMS

[Received by the International Bureau on 2 July 1991 (02.07.91); original claims 1-28 replaced by amended claims 1-26 (7 pages)]

1. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins produced sequentially by secretion from Staphylococcus aureus during growth phase, isolation from growth media by precipitation or centrifugation, and purification by chromatographic techniques.

2. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, said enterotoxins obtained by isolation from high enterotoxin producing mutant strains of Staphylococcus aureus.

3. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxin fragments, said fragments obtained by isolation from high enterotoxin producing mutant strains of Staphylococcus aureus.

4. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, said enterotoxins obtained by isolation from high enterotoxin producing bacteria capable of expressing Staphylococcus aureus enterotoxin genes.

5. A method of treating cancer in a patient which comprises the single step of administering to

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the patient a tumoricidally effective amount of Staphylococcal enterotoxin fragments, said fragments obtained by isolation from high enterotoxin producing bacteria capable of expressing Staphylococcus aureus enterotoxin genes.

6. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, said enterotoxins obtained by isolation from high enterotoxin producing cells capable of expressing Staphylococcus aureus enterotoxin genes.

7. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxin fragments, said fragments obtained by isolation from high enterotoxin producing cells capable of expressing Staphylococcus aureus enterotoxin genes.

8. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins, said enterotoxins obtained by isolation from Staphylococcus aureus which has been treated with chemical mutagens to obtain high enterotoxin producing strains.

9. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins fragments, said fragments obtained by isolation from Staphylococcus aureus

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which has been treated with chemical mutagens to obtain high enterotoxin producing strains.

10. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcus enterotoxins which have been chemically derivatized to minimize or to delete emetic and skin sensitizing toxicity while retaining T cell and cytokine stimulating activity.

11. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins utilizing single or multiple injections.

12. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of Staphylococcal enterotoxins incorporated in an adjuvant selected from the group consisting of aluminum hydroxide, liposomes, water in oil emulsions, pluronic triblock polymers, and saponin.

13. A method of treating cancer in a patient which comprises the single step of administering to a host a tumoricidally effective amount of Staphylococcal enterotoxin bound to solid supports to stimulate and to enrich a T lymphocyte population in vitro with antitumor activity for subsequent administration to a tumor-bearing patient.

14. A method of treating cancer in a patient which comprises administering to a host a

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tumorcidally effective amount of Staphylococcal enterotoxin used in combination with cytokines selected from the group consisting of interferon and tumor necrosis factor, in vitro to induce augmented T lymphocyte proliferation and antitumor activity for subsequent administration to a tumor-bearing patient.

15. A method of simultaneously activating cytokine mediators selected from the group consisting of interleukin 1, interleukin 2, tumor necrosis factor and interferon, and procoagulant systems in a host which comprises the single step of administering to the host a tumorcidally effective amount of Staphylococcal enterotoxins.

16. A method of simultaneously augmenting natural killer cell cytotoxicity, activating cytokine mediators selected from the group consisting of interleukin 1, interleukin 2, tumor necrosis factor, interferon, activating procoagulant systems, and stimulating T-lymphocyte mitogenicity, serine protease and thromboglobulin production in a host comprising a single step of administering to the host a tumorcidally effective amount of Staphylococcal enterotoxins.

17. The method of Claims 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15, or 16 in which the tumorcidally effective amount of Staphylococcal enterotoxin is between 0.5 μ g (enterotoxin) / kg (body weight) and 150 μ g (enterotoxin) / kg (body weight).

18. The method of Claims 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 11, 12, 13, 14, 15 or 16 further comprising

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the step of administering a toxicity attenuating dosage of a compound which is a non-steroidal, anti-inflammatory agent, and cyclooxygenase and prostaglandin synthesis inhibitor, to said patient.

19. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumorcidally effective amount of Streptococcus pyrogenic exotoxin.

20. A method of simultaneously activating cytokine mediators selected from the group consisting of interleukin 1, interleukin 2, tumor necrosis factor and interferon, and procoagulant systems in a host which comprises the single step of administering to the host a tumorcidally effective amount of Streptococcus pyrogenic exotoxin.

21. The method of Claim 19 or 20 in which the tumorcidally effective amount of Streptococcus pyrogenic exotoxin is between 2 μ g (exotoxin) / kg (body weight) and 150 μ g (exotoxin) / kg (body weight).

22. The method of Claims 19 or 20 further comprising the step of administering a toxicity attenuating dosage of a compound which is non-steroidal, anti-inflammatory agent, and cyclooxygenase and prostaglandin synthesis inhibitor, to said patient.

23. A method of treating cancer in a patient which comprises the single step of administering to

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the patient a tumoricidally effective amount of a compound whose molecules are functionally and structurally similar to Staphylococcal enterotoxins, and which have the properties of superantigens, binding to both TCR-CD4 complex of T lymphocytes via the V_{β} domain and binding to class II MHC molecules (IE or IA bearing) on antigen presenting cells wherein a complex is formed that is highly stimulatory for T cells.

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transformed cell with surface expression of minor lymphocyte stimulating locus and consequent potent activation and proliferation of T lymphocytes with V_{β} receptors and anti-tumor responses.

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24. A method of inducing a tumoricidal response in a patient with active compounds selected from the group consisting of enterotoxins A, B, C, D, E and F, pyrogenic exotoxins and biologically active fragments of enterotoxins said method comprising the steps of attaching said active compound molecules to antigen presenting cells in a host to form a complex; incubating said complex with T cell populations in said host to obtain activated T cell populations; and infusing said activated T cell population into a patient.

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25. A compound useful in tumoricidal therapy consisting of a synthetic polypeptide characterized by substantial structural homology to Staphylococcal enterotoxins and streptococcal pyrogenic exotoxins, wherein said homology includes statistically significant sequence homology, alignment of cysteine residues, and similar hydrophathy profiles.

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26. A method of treating cancer in a patient which comprises the single step of administering to the patient a tumoricidally effective amount of tumor cells which have been transfected with an enterotoxin gene, said tumor cell administration resulting in a

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STN	SEC1	SEC2	SEC3	SEC4	SEC5	SEC6	SEC7	SEC8	SEC9	SEC10	SEC11	SEC12	SEC13	SEC14	SEC15	SEC16	SEC17	SEC18	SEC19	SEC20	SEC21	SEC22	SEC23	SEC24	SEC25	SEC26	SEC27	SEC28	SEC29	SEC30	SEC31	SEC32	SEC33	SEC34	SEC35	SEC36	SEC37	SEC38	SEC39	SEC40	SEC41	SEC42	SEC43	SEC44	SEC45	SEC46	SEC47	SEC48	SEC49	SEC50	SEC51	SEC52	SEC53	SEC54	SEC55	SEC56	SEC57	SEC58	SEC59	SEC60	SEC61	SEC62	SEC63	SEC64	SEC65	SEC66	SEC67	SEC68	SEC69	SEC70	SEC71	SEC72	SEC73	SEC74	SEC75	SEC76	SEC77	SEC78	SEC79	SEC80	SEC81	SEC82	SEC83	SEC84	SEC85	SEC86	SEC87	SEC88	SEC89	SEC90	SEC91	SEC92	SEC93	SEC94	SEC95	SEC96	SEC97	SEC98	SEC99	SEC100	SEC101	SEC102	SEC103	SEC104	SEC105	SEC106	SEC107	SEC108	SEC109	SEC110	SEC111	SEC112	SEC113	SEC114	SEC115	SEC116	SEC117	SEC118	SEC119	SEC120	SEC121	SEC122	SEC123	SEC124	SEC125	SEC126	SEC127	SEC128	SEC129	SEC130	SEC131	SEC132	SEC133	SEC134	SEC135	SEC136	SEC137	SEC138	SEC139	SEC140	SEC141	SEC142	SEC143	SEC144	SEC145	SEC146	SEC147	SEC148	SEC149	SEC150	SEC151	SEC152	SEC153	SEC154	SEC155	SEC156	SEC157	SEC158	SEC159	SEC160	SEC161	SEC162	SEC163	SEC164	SEC165	SEC166	SEC167	SEC168	SEC169	SEC170	SEC171	SEC172	SEC173	SEC174	SEC175	SEC176	SEC177	SEC178	SEC179	SEC180	SEC181	SEC182	SEC183	SEC184	SEC185	SEC186	SEC187	SEC188	SEC189	SEC190	SEC191	SEC192	SEC193	SEC194	SEC195	SEC196	SEC197	SEC198	SEC199	SEC200	SEC201	SEC202	SEC203	SEC204	SEC205	SEC206	SEC207	SEC208	SEC209	SEC210	SEC211	SEC212	SEC213	SEC214	SEC215	SEC216	SEC217	SEC218	SEC219	SEC220	SEC221	SEC222	SEC223	SEC224	SEC225	SEC226	SEC227	SEC228	SEC229	SEC230	SEC231	SEC232	SEC233	SEC234	SEC235	SEC236	SEC237	SEC238	SEC239	SEC240	SEC241	SEC242	SEC243	SEC244	SEC245	SEC246	SEC247	SEC248	SEC249	SEC250	SEC251	SEC252	SEC253	SEC254	SEC255	SEC256	SEC257	SEC258	SEC259	SEC260	SEC261	SEC262	SEC263	SEC264	SEC265	SEC266	SEC267	SEC268	SEC269	SEC270	SEC271	SEC272	SEC273	SEC274	SEC275	SEC276	SEC277	SEC278	SEC279	SEC280	SEC281	SEC282	SEC283	SEC284	SEC285	SEC286	SEC287	SEC288	SEC289	SEC290	SEC291	SEC292	SEC293	SEC294	SEC295	SEC296	SEC297	SEC298	SEC299	SEC300	SEC301	SEC302	SEC303	SEC304	SEC305	SEC306	SEC307	SEC308	SEC309	SEC310	SEC311	SEC312	SEC313	SEC314	SEC315	SEC316	SEC317	SEC318	SEC319	SEC320	SEC321	SEC322	SEC323	SEC324	SEC325	SEC326	SEC327	SEC328	SEC329	SEC330	SEC331	SEC332	SEC333	SEC334	SEC335	SEC336	SEC337	SEC338	SEC339	SEC340	SEC341	SEC342	SEC343	SEC344	SEC345	SEC346	SEC347	SEC348	SEC349	SEC350	SEC351	SEC352	SEC353	SEC354	SEC355	SEC356	SEC357	SEC358	SEC359	SEC360	SEC361	SEC362	SEC363	SEC364	SEC365	SEC366	SEC367	SEC368	SEC369	SEC370	SEC371	SEC372	SEC373	SEC374	SEC375	SEC376	SEC377	SEC378	SEC379	SEC380	SEC381	SEC382	SEC383	SEC384	SEC385	SEC386	SEC387	SEC388	SEC389	SEC390	SEC391	SEC392	SEC393	SEC394	SEC395	SEC396	SEC397	SEC398	SEC399	SEC400	SEC401	SEC402	SEC403	SEC404	SEC405	SEC406	SEC407	SEC408	SEC409	SEC410	SEC411	SEC412	SEC413	SEC414	SEC415	SEC416	SEC417	SEC418	SEC4
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FIGURE 1

PICTURE 1 (continued)

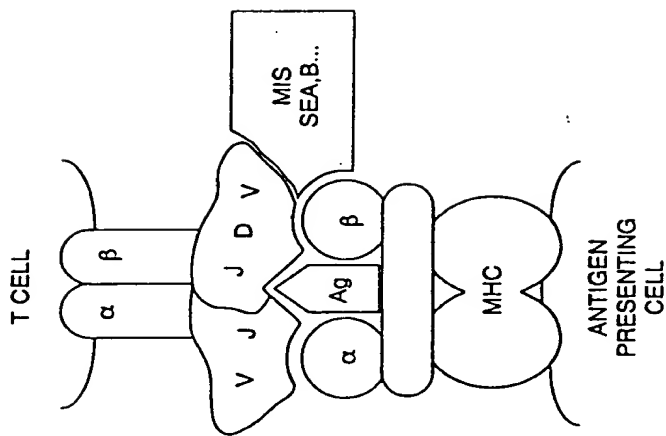


FIG. 3

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SUBSTITUTE SHEET

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)

Category	Citation of Document with indication, where appropriate, of the relevant passages	Relevant to Claim 1
X	Biological Abstracts, volume 84, no. 5 issued 01 September 1987. Shcheglovitova et al., "Nonspecific antitumor activity of staphylococcal enterotoxins". See Abstract 48345. EKSP ONKOL, 9(1): 28-30.	1-12.15.23 24.25.27
X	Infection and Immunity, vol. 57, no. 7, issued July 1989. Penarrubia et al., "Selective Proliferation of Natural Killer Cells among Monoate Depleted Peripheral Blood Mononuclear Cells as a Result of Stimulation with Staphylococcal Enterotoxin B" pgs. 2057-2065. See "introduction".	1-12.15.23 25.27
Y	Proc. Natl. Acad. Sci. vol. 72, no. 9, issued September 1975. Carswell et al., "An endotoxin-induced serum factor that causes necrosis of tumors" pgs. 3666-3670. See entire document.	1-15.17.18 22-27
Y	WO/A,88/02632 (Goldberg et al.) 21 April 1988. See Abstract.	16.19-21
Y	Infection and Immunity, vol. 57, no. 1, issued January 1989. Fast et al. "Toxic Shock Syndrome Associated Staphylococcal and Streptococcal Pyrogenic Toxins are Potent Inducers of Tumor Necrosis Factor Production," pgs. 291-294. See Abstract.	1-15.17.18 22-27

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